Prognostic Value of Mitotic Counts in Breast Cancer of Saudi Arabian Patients

ABDELBASET BUHMEIDA¹, JAUDAH AL-MAGHRABI², ADNAN MERDAD³, FATIMA AL-THUBAITY³, ADEEL CHAUDHARY¹, MAMDOOH GARI¹, ADEL ABUZENADAH¹, YRJÖ COLLAN⁴, KARI SYRJÄNEN⁵ and MOHAMMED AL-QAHTANI¹

¹Center of Excellence in Genomic Medicine Research, and Departments of ²Pathology and ³Surgery, King Abdul-Aziz University, Jeddah, Saudi Arabia; Departments of ⁴Pathology, and ⁵Oncology and Radiotherapy, Turku University Hospital, Turku, Finland

Abstract. *Background: Ouantitative methods* in combination with other objective prognostic criteria can improve the evaluation of a cancer patient's prognosis, and possibly predict response to therapy. One of the important prognostic and predictive markers is the mitotic count, which has proven valuable in many aspects. In this study, the prognostic value of the mitotic count was assessed in breast cancer (BC) patients in Saudi Arabia. Patients and Methods: The study comprised a series of 87 patients diagnosed and treated for breast cancer at the Departments of Surgery and Oncology, King Abdul-Aziz University Hospital, between 2000 and 2008. Mitotic counts were carried out using a standard laboratory microscope (objective, $\times 40$; field diameter, 420 µm). The number of mitotic figures in 10 consecutive high-power fields (hpf) from the most cellular area of the sample gave the mitotic activity index (MAI, mitotic figures/10 hpf). The standardized mitotic index (SMI) recorded the mitotic count as the number of mitotic figures by area of the neoplastic tissue in the microscopic field, thus the number of mitoses in 10 consecutive fields was corrected for the volume fraction and field size (mitotic figures/mm²). Results: The means of MAI and SMI of the tumors in the entire series of 87 patients were 15 mitotic figures/10hpf (range 4-45) and 4 mitotic figures/mm² (range 1-9), respectively. The mitotic counts were higher in advanced stages than in early cancer (p<0.04). The mitotic counts were significantly larger in patients with high-grade tumor (p<0.004) and in cases with

Correspondence to: Dr. Abdelbaset Buhmeida, MD, Ph.D., Center of Excellence in Genomic Medicine Research (CEGMR), King Abdul-Aziz University, P.O. Box: 80216 Jeddah 21589, Saudi Arabia. Tel: +9662 6401000 ext: 25481, Fax: +9662 6952521, e-mail: abuhmeida@kau.edu.sa, abuhme@utu.fi

Key Words: Mitotic counts, breast cancer, prognosis.

tumor metastasis (p<0.004). The mitotic counts were also significantly larger in the recurrent cases than in nonrecurrent ones (p<0.02). Conclusion: The quantitatively measurable mitotic counts of cancer cell nuclei are of significant prognostic value in invasive ductal carcinoma of the breast in Saudi Arabia and the mean cut-off values of MAI and SMI can be applied as objective (quantitative) criteria to distinguish breast cancer patients into groups with favorable and less favorable prognosis.

According to the Saudi Cancer Registry Report (2005) (1), breast cancer (BC) is the most common cancer among women, accounting for 22.4% of all newly diagnosed carcinomas, with an age-specific incidence rate of 15.4/100,000. The median age at diagnosis is 47 years (range 18-96 years). BC has been intensely studied worldwide, but many aspects still remain unclear, including some intriguing special features of BC encountered in different global regions. The possibility that these geographic differences may have a genetic basis is one favored hypothesis. The variation in the distribution of different BC genetic marker haplotypes with a clear difference in distribution between Western Central Africa and Northern Africa and similarly between Asia and Europe has substantiated this suggestion (2).

Approximately, 20%-30% of the patients with lymph node-negative (LN^-) BC die of recurrent disease. A relative survival improvement of 15%-20% over the next decade might be expected from improvements in adjuvant systemic therapy (3). Accurate and reliable prognostic markers are needed to help identify the high-risk patients. The BC prognosis can be evaluated by combining different clinico-pathological features such as tumor size, stage, grade and LN status (4). Also the histological grading system provides high prognostic potential (5, 6), but it suffers from being subjective and still leaves a substantial group of patients with an unclear prognosis (7). In this respect, accurate quantitative measurements would be expected to be more reproducible than the subjective methods of tumor grading (8).

Many recent studies have shown that proliferation markers exceed the prognostic value of classical predictors (9-12). A variety of methods have become available to assess the rate of proliferation based on the cell cycle (13, 14). The growth fraction can be evaluated using immunohistochemistry for different proliferation-associated antigens, such as Ki-67 (15), topoisomerase IIa (16), proliferating cell nuclear antigen (PCNA) (17) and geminin (14), or by analysis of the S-phase fraction using DNA flow cytometry or DNA static image cytometry (18). However, the S-phase fraction method is hampered by pronounced intra-tumor heterogeneity (19). Therefore, mitosis counting and the Ki-67 index are the most practical methods. However, in a Finnish material it became quite clear that the mitotic count was even much better prognosticator than quantified Ki-67 staining (quantified as the fraction of positively staining nuclei) (20). Out of these two methods, mitosis counting has been best studied from a methodological point of view, based on larger retrospective and prospective studies (21). Mitotic counting has been reported to be a powerful, practical, easily assessable, inexpensive and highly reproducible prognosticator (9, 21-24). Furthermore, several studies have indicated that the mitotic count is the most important constituent of the histological grade (25, 26), but well-known problems with reproducibility of the grading exist due to the lack of strict protocols (27, 28). Aaltomaa et al. (29) have suggested that all types of BC could be graded using the same principles when mitotic indices are determined, based on the observed minor differences only in the proliferative activity between ductal carcinomas and all special BC forms (29).

Among a wide range of quantitative histopathology approaches for unbiased assessment of potential prognostic factors, nuclear morphometry (30-32) and mitotic counting (33), have been shown to be able to distinguish between benign and malignant lesions. With others, we have suggested that the mitotic count in combination with other objective prognostic criteria could improve the evaluation of prognosis in BC and possibly predict response to therapy, independently of the geographic peculiarities of this disease.

As part of our efforts to introduce a mitotic count grading system specifically suitable for BC in Saudi women, the prognostic value of mitotic count was assessed in BC patients in Saudi Arabia, with special reference to similar data reported from other countries.

Patients and Methods

The material for this study was derived from a cohort of 201 consecutive women diagnosed with BC at the Department of Pathology, King Abdul-Aziz University, Jeddah, Saudi Arabia

between 2000 and 2008. Patients were excluded from this study on the basis of the following criteria: histopathological diagnosis was not invasive ductal carcinoma (IDC); patient history, medical files or BC specimens were not found. The remaining cohort of 87 women with IDC was eligible for the counting of mitotic figures.

The pertinent clinicopathological features (age, menopausal status, stage, grade, and LNN status) and follow-up and survival data were collected from the patient files and are summarized in Table I. The mean age at the time of diagnosis was 47.5 years (range: 19-81 years).

Treatment and follow-up. Almost all the patients were subjected to surgery, *i.e.*, lumpectomy, radical or modified radical mastectomy with axillary clearance. Postoperative early adjuvant systemic therapy in the form of chemotherapy, radiotherapy and hormonal therapy was given to 72%, 56% and 38% of the patients, respectively. After treatment, the patients were seen at 6-12 month intervals until death or the end of follow-up (FU) in mid August, 2009. Some patients were lost to FU. The mean FU time for the whole series was 47 months (range: 4-118 month). During FU, the patients were subjected to repeated clinical examination and bone isotope scan, chest and abdominal-pelvic CAT scan were performed whenever needed. In most instances, the cause of death was obvious on clinical grounds alone. Autopsy was not performed in any case.

During the FU period, 15 (17%) patients developed recurrence and 12 (13%) patients developed distant metastasis in different organs. Disease-free survival (DFS) and disease-specific survival (DSS) were calculated as the time from diagnosis to the appearance of recurrent disease (or date last seen disease-free), and time from diagnosis to death (due to disease) or to the date last seen alive, respectively. In calculating the DSS, the patients who died of other or unknown causes were censored.

Counting mitotic figures. All the tissue samples has been obtained from the primary tumor at the time of diagnosis. The samples were fixed in buffered formalin and embedded in paraffin. Sections were cut at 5 µm and stained with H&E. Mitotic figures were characterized by an absent nuclear membrane with clear, hairy extensions of nuclear material (condensed chromosomes) either clumped (beginning metaphase), in a plane (metaphase/anaphase) or in separate chromosomal aggregates (anaphase/telophase). The cytoplasm of the mitotic cells was often larger during mitosis than in the resting cells. Special attention was paid to distinguishing between apoptotic bodies and mitotic figures (34). The recognition of at least one chromosome, usually appearing as a small protuberance at the outline of the chromosome clump was required for inclusion in the mitotic count. The absence of nuclear membrane was also an important feature, but did not alone constitute mitosis.

Sampling rules. The mitotic figures were counted in the most cellular area at the periphery of the tumor from 10 consecutive high- power fields (hpf) (35). Necrotic and inflammatory areas were avoided. If several areas met these criteria, the area with the highest number of mitotic figures, assessed subjectively, was chosen. Two parallel clearly separate chromosome clumps were counted as one mitotic figure. Mitotic counting was carried out using a standard laboratory microscope (objective, \times 40; field diameter, 440 µm). Two methods were used record mitoses in the

Table I	. Clinico	pathological	characteristics	of the	patients.

Characteristic	No. of patients (%)
Age (years)	
<50	50 (57%)
>50	37 (43%)
Menopausal status	
Premenopausal	48 (55%)
Postmenopausal	39 (45%)
Localization	
Right	43 (49%)
Left	44 (51%)
Neurovascular invasion	
No	25 (29%)
Yes	37 (42%)
Unknown	25 (29%)
Lymph node	
NO	24 (28%)
N1	33 (38%)
Nx	30 (34%)
Metastasis	
M0	38 (44%)
M1	12 (13%)
Mx	37 (43%)
Grade	
G1	17 (20%)
G2	48 (55%)
G3	21 (24%)
Gx	1 (1%)
Stage	
1	16 (18%)
2	44 (51%)
3	4 (5%)
4	12 (14%)
Unknown x	11 (12%)
Recurrence during follow-up	
Yes	15 (17%)
No	56 (65%)
Unknown	16 (18%)
Response to treatment	
CR	45 (51%)
PR	7 (8%)
PD	11 (13%)
Unknown	24 (28%)
Status at the end of follow-up	
Alive	66 (76%)
Died of disease	8 (9%)
Unknown	13 (15%)

CR: complete response, PR: partial response, PD: progressive disease.

cancer cells the mitotic activity index (MAI) and the standardized mitotic index (SMI). The number of mitotic figures in the 10 consecutive hpf gave the mitotic activity index (MAI). From the counted areas, SMI (also called M/Vv index) was calculated as the number of mitotic figures by area of the neoplastic tissue in the microscopic field. Thus the number of mitoses in 10 consecutive fields was corrected for the volume fraction and field size (37). SMI=k $(\Sigma MI)/(\Sigma Vv)$ Table II. Clinicopathological features and associated mitotic activity (MAI) and standardized mitotic (SMI) indices.

	Mean mitotic count <i>p</i> -value		
Clinicopathological features	MAI	SMI	
Age	0.840	0.40	
Menopausal status	0.790	0.84	
Site (L, R)	0.770	0.02	
Margins	0.760	0.54	
Invasion	0.870	0.85	
Lymph node	0.690	0.47	
Metastasis	0.004	0.01	
Grade	0.004	0.01	
Stage	0.040	0.05	
Response to treatment	0.050	0.01	
Recurrence	0.020	0.01	
DFS	0.300	0.30	
DSS	0.300	0.40	

L: Left, R: right, DFS: disease-free survival, DSS: disease-specific survival.

Where $k=100/r^2$, r=the radius of the field and MI=number of mitotic figures in the studied fields. Vv is the volume fraction of malignant epithelium in the field.

Statistical analysis. Statistical analyses were performed using the SPSS® (SPSS, Inc., Chicago state, USA) and STATA (Stata Corp., town, TX, USA) software packages (PASW Statistics for Windows, version 18.0.1 and STATA/SE 11.0). Student t-tests and ANOVA were used to test differences between the groups. Bivariate correlations between the mitotic counts and DFS and DSS were evaluated using Pearson's correlation test. For univariate survival analysis, Kaplan-Meier curves were plotted and differences between the strata (MAI and SMI cut-offs) were analyzed using the log-rank test. In addition, multivariate analysis was performed using Cox's regression model with known prognostic predictors (age, family history, site, tumor grade, LNN involvement, response to treatment, stage) were entered in stepwise backward approach, to evaluate the independent prognostic value of MAI and SMI. In all the analyses, p-values below 0.05 were regarded as significant.

Results

Clinicopathological features. The correlation of the mitotic counts (MAI and SMI) with the different clinicopathological features is shown in Table II. MAI and SMI means were 15 mitotic figures/10 hpf, (range 4-45), and 4 mitotic figures/mm² (range 1-9), respectively, in the whole series of 87 samples. The MAI and SMI means were used as the cutoff in further calculations to correlate the mitotic counts with the clinical parameters and disease outcome.

Higher values of SMI were seen in the left breast tumors than the right side tumors (p < 0.02), while MAI did not show this trend (p < 0.77). Significant associations were observed between mitotic count and histological grade.



Figure 1. MAI as predictor of disease-free survival (DFS).

High-grade tumors showed higher mitotic counts (19 mitotic figures/10 hpf) for MAI and for SMI (5.1 mitotic figures/mm²) as compared with low-grade tumors (12 mitotic figures/10 hpf and 3.8 mitotic figures/mm², respectively), (p<0.0004, MAI; p<0.01, SMI, respectively). Similarly, mitotic counts were significantly higher in the tumors that subsequently recurred (19 mitotic figures/10 hpf for MAI and 5 mitotic figures/mm² for SMI) when compared with the non-recurrent ones (13.9 mitotic figures/10 hpf and 3.9 mitotic figures/mm²; p < 0.02, p < 0.01, respectively). In the same way, mitotic counts were higher in the patients who developed metastasis (21 mitotic figures/10 hpf and 5.3 mitotic figures/mm²) than in those who did not by the end of the follow-up (13 mitotic figures/10 hpf and 3.9 mitotic figures/mm²; p < 0.0004, MAI; p < 0.01, SMI, respectively). The mitotic counts were also higher in the advanced stages (21 mitotic figures/10hpf and 5.4 mitotic figures/mm²) than in early stages (15 mitotic figures/10 hpf and 4 mitotic figures/mm²) (p<0.04, MAI; p < 0.05, SMI, respectively). There was also a significant association between mitotic count and response to treatment: the mean mitotic counts of patients with complete response (CR), partial response (PR) and progressive disease (PD) were 13 mitotic figures/10 hpf for MAI and 4 mitotic figures/mm² for SMI, 16 mitotic figures/10 hpf for MAI and 4.9 mitotic figures/mm² for SMI, and 19 mitotic figures/10 hpf for MAI and 5 mitotic figures/mm² for SMI (*p*<0.05, MAI; *p*<0.01, SMI), respectively.

The values of MAI and SMI were slightly higher in the LN^+ patients than in LN^- patients (p<0.6, MAI; p<0.4, SMI, respectively). The same trend was observed between MAI/SMI and disease outcome, both being higher among the



Figure 2. SMI as predictor of disease-specific survival (DSS).

women who died of their disease as compared with those who were alive, although the difference did not reach significance (p<0.16, MAI; p<0.10, SMI, respectively). In contrast, there was no relationship between age and mitotic count, which was identical in the patients below and above the mean age of 47.5 years (p<0.84, MAI; p<0.40, SMI, respectively). Similarly, the mitotic count was associated with neither the involvement of the tumor margins (p<0.76, MAI; p<0.54, SMI, respectively) nor with tumor invasion to blood vessels or nerves (p<0.87, MAI; p<0.85, SMI, respectively).

Survival analysis. In the univariate (Kaplan-Meier) survival analysis, MAI (with mean as the cut-off) showed a trend towards being a predictor of DFS (log-rank p<0.3) (Figure 1). At 6 years, 20% of the patients with lower MAI showed recurrence, as compared to 33% of the patients with higher MAI (Figure 1). Mitotic count did not show any significant correlation with DSS (Figure 2, p<0.4).

Out of the variables entered in the multivariate regression model, response to therapy was the only independent predictor of DFS, with HR=3.42 (95% CI 1.77-6.61) for women with CR to be recurrence-free as compared to those with PR or PD. As with DFS, MAI or SMI were not independent predictors of DSS in the multivariate model, where none of the other variables proved to be independent predictors.

Discussion

A close correlation between the mitotic count and some of the clinicopathological features and also the disease outcome was shown in the BC patients. However, the biological mechanisms responsible for these mitotic count variations in the tumor cells remain to be disclosed, although certain mutations in growth-regulating genes may contribute to the high mitotic activity seen (37). The significant factors observed in this study reflect the prognostic variables in the early stages of follow-up in Saudi BC. These mean values MAI and SMI were useful in separating the patients with favorable and unfavorable outcome of the disease in the present cohort. The proliferative activity of the Saudi material was within the ranges reported in a Finnish BC series where the corresponding values were 10.7 mitotic figures/10 hpf and 13.8 mitotic figures/mm² and in other European studies as well (38). However, these figures were much lower than in a Nigerian and African-American BC series (39). The Finnish premenopausal patients (40) had higher values of the proliferative indices than the postmenopausal patients which was in contrast to the Nigerian patients studied by Ikpatt et al. (39) where the mitotic count was higher in the postmenopausal patients. However, no statistically significant difference between the menopausal statuses was shown in the present study.

The different mitotic counts observed in the present series might reflect actual biological differences between BC in these populations. It is well known that significantly different tumor cell populations, clones, with dissimilar biology, exist in highly proliferating advanced BC. These different clones may have different *p53* status, DNA ploidy, proliferation rates and nuclear morphology (41).

The observation that tumors with higher mitotic counts were associated with the presence of LN metastasis was similar to other studies and requires further assessment. It would seem feasible that tumors with higher mitotic counts are more aggressive and more likely to be associated with LN involvement at diagnosis. In accordance with other similar cohorts (39, 42), the present study showed that the mitotic counts were correlated with the tumor grade and stage, high-grade tumors showing higher mitotic counts, which would be expected as the mitotic count is considered the most important constituent of the histological grade in individual tumors (21).

In the present study, a significant association was shown between the mitotic counts and the response to treatment: an objective response to adjuvant systemic therapy was observed among the patients with lower mitotic activity, in contrast to those with a higher proliferation rate who developed PD. This was in contrast to some other studies which showed that patients with rapidly proliferating tumors benefited from adjuvant systemic therapy more than patients with low proliferation rates (43, 44). The reasons for this discrepancy remain obscure at the moment.

In the present series, the mitotic count proved to be of some use in discriminating between patients with poor and favorable DFS in the univariate survival analysis with the patients with higher mitotic counts showing a higher rate of recurrence compared with those showing lower mitotic activity at baseline, although the difference did not reach statistical significance. As seen in Figures 1 and 2, the difference became more evident after mid- to long-term FU, and the full importance of these data merits confirmation in a larger series with extended (>10 years) FU. In the multivariate survival analysis, however, neither MAI nor SMI proved to be of any value as independent predictors of DFS or DSS.

In conclusion, increased cell proliferation in BC in Saudi Arabian patients correlates strongly with several indicators of poor prognosis, and the mean cut-off values of MAI and SMI can be applied as objective (quantitative) criteria to distinguish between BC patients with favorable or less favorable prognosis.

References

- 1 Al-Eid H and Arteh S: Cancer Incidence Report Saudi Arabia. pp. 1-99, 2005.
- 2 Buyru N, Altinisik J, Demokan S and Dalay N: *p53* genotypes and haplotypes associated with risk of breast cancer. Cancer Detect Prev 31: 207-213, 2007.
- 3 Group E. B. C. T. C: Effects of chemotherapy and hormonal therapy for early breast cancer on recurrence and 15-year survival: an overview of the randomised trials. Lancet *365*: 1687-1717, 2005.
- 4 McBride R, Hershman D, Tsai WY, Jacobson JS, Grann V and Neugut A: I. Within-stage racial differences in tumor size and number of positive lymph nodes in women with breast cancer. Cancer 110: 1201-1208, 2007.
- 5 Collan Y, Kuopio T and Alanen K: Scope and concepts of quantitive histopathology. Acta Stereol *11(1)*: 3-23, 1992.
- 6 Sotiriou C, Loi S, Harris A, Fox S, Smeds J et al: Gene expression profiling in breast cancer: understanding the molecular basis of histologic grade to improve prognosis. J Natl Cancer Inst 98: 262-272, 2006.
- 7 Page DL: Prognosis and breast cancer. Recognition of lethal and favorable prognostic types. Am J Surg Pathol *15*: 334-349, 1991.
- 8 Frierson HF Jr., Wolber RA, Berean KW, Franquemont DW, Gaffey MJ, Boyd JC and Wilbur DC: Interobserver reproducibility of the Nottingham modification of the Bloom and Richardson histologic grading scheme for infiltrating ductal carcinoma. Am J Clin Pathol 103: 195-198, 1995.
- 9 Groenendijk RP, Bult P, Noppen CM, Boetes C, Ruers TJ and Wobbes T: Mitotic activity index in interval breast cancers. Eur J Surg Oncol 29: 29-31, 2003.
- 10 Manders P, Bult P, Sweep CG, Tjan-Heijnen VC and Beex LV: The prognostic value of the mitotic activity index in patients with primary breast cancer who were not treated with adjuvant systemic therapy. Breast Cancer Res Treat 77: 77-84, 2003.
- 11 Volpi A, Bacci F, Paradiso A, Saragoni L, Scarpi E, Ricci M, Aldi M, Bianchi S, Muretto P, Nuzzo F, Simone G, Mangia A, Schittulli F and Amadori D: Prognostic relevance of histological grade and its components in node-negative breast cancer patients. Mod Pathol 17: 1038-1044, 2004.

- 12 Meyer JS, Alvarez C, Milikowski C, Olson N, Russo I, Russo J, Glass A, Zehnbauer BA, Lister K and Parwaresch R: Breast carcinoma malignancy grading by Bloom-Richardson system vs. proliferation index: reproducibility of grade and advantages of proliferation index. Mod Pathol 18: 1067-1078, 2005.
- 13 Daidone MG and Silvestrini R: Prognostic and predictive role of proliferation indices in adjuvant therapy of breast cancer. J Natl Cancer Inst Monogr 30: 27-35, 2001.
- 14 Sherbet GV and Patil D: Genetic abnormalities of cell proliferation, invasion and metastasis, with special reference to gynaecological cancers. Anticancer Res 23: 1357-1371, 2003.
- 15 Karanikas G, Koronakis N, Lagoudianakis EE, Grosomanidis D, Karavitis G, Koukoutsis I, Pappas A, Kotzadimitriou K, Papadima A, Chrysikos J, Zografos G, Xepapadakis G and Manouras A: The value of proliferation indexes in breast cancer. Eur J Gynaecol Oncol 31: 181-184, 2010.
- 16 Nakopoulou L, Lazaris AC, Kavantzas N, Alexandrou P, Athanassiadou P, Keramopoulos A and Davaris P: DNA topoisomerase II-alpha immunoreactivity as a marker of tumor aggressiveness in invasive breast cancer. Pathobiology 68: 137-143, 2000.
- 17 Stuart-Harris R, Caldas C, Pinder SE and Pharoah P: Proliferation markers and survival in early breast cancer: a systematic review and meta-analysis of 85 studies in 32,825 patients. Breast *17*: 323-334, 2008.
- 18 Moureau-Zabotto L, Bouchet C, Cesari D, Uzan S, Lefranc JP, Antoine M, Genestie C, Deniaud-Alexandre E, Bernaudin JF, Touboul E and Fleury-Feith J: Combined flow cytometry determination of S-phase fraction and DNA ploidy is an independent prognostic factor in node-negative invasive breast carcinoma: analysis of a series of 271 patients with stage I and II breast cancer. Breast Cancer Res Treat 91: 61-71, 2005.
- 19 Bergers E, van Diest PJ and Baak JP: Tumour heterogeneity of DNA cell cycle variables in breast cancer measured by flow cytometry. J Clin Pathol 49: 931-937, 1996.
- 20 Jalava P, Kuopio T, Juntti-Patinen L, Kotkansalo T, Kronqvist P and Collan Y: Ki-67 immunohistochemistry: a valuable marker in prognostication but with a risk of misclassification: proliferation subgroups formed based on Ki-67 immunoreactivity and standardized mitotic index. Histopathology 48: 674-682, 2006.
- 21 van Diest PJ, van der Wall E and Baak JP: Prognostic value of proliferation in invasive breast cancer: a review. J Clin Pathol 57: 675-681, 2004.
- 22 Montironi R, Collan Y, Scarpelli M, Sisti S, Barbatelli G, Carnevali A, Pisani E and Mariuzzi GM: Reproducibility of mitotic counts and identification of mitotic figures in malignant glial tumors. Appl Pathol 6: 258-265, 1988.
- 23 Vesalainen S, Lipponen P, Talja M and Syrjanen K: Mitotic activity and prognosis in prostatic adenocarcinoma. Prostate 26: 80-86, 1995.
- 24 Baak JP, van Diest PJ, Voorhorst FJ, van der Wall E, Beex LV, Vermorken JB, Janssen EA and Gudlaugsson E: The prognostic value of proliferation in lymph-node-negative breast cancer patients is age dependent. Eur J Cancer 43: 527-535, 2007.
- 25 Genestie C, Zafrani B, Asselain B, Fourquet A, Rozan S, Validire P, Vincent-Salomon A and Sastre-Garau X: Comparison of the prognostic value of Scarff-Bloom-Richardson and Nottingham histological grades in a series of 825 cases of breast cancer: major importance of the mitotic count as a component of both grading systems. Anticancer Res 18: 571-576, 1998.

- 26 Le Doussal V, Tubiana-Hulin M, Friedman S, Hacene K, Spyratos F and Brunet M: Prognostic value of histologic grade nuclear components of Scarff-Bloom-Richardson (SBR). An improved score modification based on a multivariate analysis of 1262 invasive ductal breast carcinomas. Cancer 64: 1914-1921, 1989.
- 27 Dalton LW, Page DL and Dupont WD: Histologic grading of breast carcinoma. A reproducibility study. Cancer 73: 2765-2770, 1994.
- 28 Boiesen P, Bendahl PO, Anagnostaki L, Domanski H, Holm E, Idvall I, Johansson S, Ljungberg O, Ringberg A, Ostberg G and Ferno M: Histologic grading in breast cancer – reproducibility between seven pathologic departments. South Sweden Breast Cancer Group. Acta Oncol 39: 41-45, 2000.
- 29 Aaltomaa S, Lipponen P, Eskelinen M, Kosma VM, Marin S, Alhava E and Syrjanen K: Mitotic indexes as prognostic predictors in female breast cancer. J Cancer Res Clin Oncol 118: 75-81, 1992.
- 30 Schondorf H and Naujoks H: Determining the nuclear area in normal breast epithelia and in the nuclei of mammary carcinomas. J Cancer Res Clin Oncol *109*: 241-244, 1985.
- 31 Axelrod DMN, Lickley H, Qian J, Christens-Barry W, Yuan Y, Fu Y and Chapman J: Effect of quantitative nuclear image features on recurrence of ductal carcinoma *in situ* (DCIS) of the breast. Cancer Inform 6: 99-109, 2008.
- 32 Buhmeida A, Al-Maghrabi J, Merdad A, Al-Thubaity F, Chaudhary A, Gari M, Abuzenadah A, Collan Y, Syrjanen K and Al-Qahtani M: Nuclear morphometry in prognostication of breast cancer in Saudi Arabian patients: comparison with European and African breast cancer. Anticancer Res 30: 2185-2191, 2010.
- 33 Kronqvist P, Kuopio T and Collan Y: Quantitative thresholds for mitotic counts in histologic grading: confirmation in nonfrozen samples of invasive ductal breast cancer. Ann Diagn Pathol 4: 65-70, 2000.
- 34 Baak JP, van Diest PJ, Ariens AT, van Beek MW, Bellot SM, Fijnheer J, van Gorp LH, Kwee WS, Los J and Peterse HC: The Multicenter Morphometric Mammary Carcinoma Project (MMMCP). A nationwide prospective study on reproducibility and prognostic power of routine quantitative assessments in The Netherlands. Pathol Res Pract 185: 664-670, 1989.
- 35 Baak JP, Van Dop H, Kurver PH and Hermans J: The value of morphometry to classic prognosticators in breast cancer. Cancer 56: 374-382, 1985.
- 36 Haapasalo H, Pesonen E and Collan Y: Volume corrected mitotic index (M/V-INDEX). The standard of mitotic activity in neoplasms. Pathol Res Pract *185*: 551-554, 1989.
- 37 Taioli E, Bradlow HL, Garbers SV, Sepkovic DW, Osborne MP, Trachman J, Ganguly S and Garte SJ: Role of estradiol metabolism and *CYP1A1* polymorphisms in breast cancer risk. Cancer Detect Prev 23: 232-237, 1999.
- 38 Baak JP, van Diest PJ, Voorhorst FJ, van der Wall E, Beex LV, Vermorken JB and Janssen EA: Prospective multicenter validation of the independent prognostic value of the mitotic activity index in lymph node-negative breast cancer patients younger than 55 years. J Clin Oncol 23: 5993-6001, 2005.
- 39 Ikpatt OF, Kuopio T and Collan Y: Proliferation in African breast cancer: biology and prognostication in nigerian breast cancer material. Mod Pathol 15: 783-789, 2002.

- 40 Kronqvist P, Kuopio T and Collan Y: Morphometric grading in breast cancer: thresholds for mitotic counts. Hum Pathol 29: 1462-1468, 1998.
- 41 Friedrich K, Dimmer V, Haroske G, Meyer W, Theissig F and Kunze KD: Correlation between p53 status, DNA ploidy, proliferation rate and nuclear morphology in breast cancer. An image cytometric study. Anal Cell Pathol 15: 85-97, 1997.
- 42 Tawfik O, Kimler BF, Davis M, Stasik C, Lai SM, Mayo MS, Fan F, Donahue JK, Damjanov I, Thomas P, Connor C, Jewell, WR, Smith H and Fabian CJ: Grading invasive ductal carcinoma of the breast: advantages of using automated proliferation index instead of mitotic count. Virchows Arch 450: 627-636, 2007.
- 43 Andre F, Khalil A, Slimane K, Massard C, Mathieu MC, Vignot S, Assi H, Delaloge S and Spielmann M: Mitotic index and benefit of adjuvant anthracycline-based chemotherapy in patients with early breast cancer. J Clin Oncol 23: 2996-3000, 2005.
- 44 Janssen EA, van Diest PJ, Soiland H, Gudlaugson E, Nysted A, Voorhorst FJ, Vermorken JB, Soreide JA and Baak JP: Success predictors of adjuvant chemotherapy in node-negative breast cancer patients under 55 years. Cell Oncol 28: 295-303, 2006.

Received August 2, 2010 Revised November 30, 2010 Accepted December 21, 2010